

Directing The Role of Histone Deacetylase Inhibitors in Cancer Therapy: A Review

Bhavini K Gharia¹, Bhanubhai N Suhagia², JagatkumarUpadhyay³

¹Department of Pharmaceutical Chemistry, Maliba Pharmacy College, Uka tarsadia university, Bardoli, India.

^{2,3}Department of Pharmaceutical Chemistry, Dharamsinh Desai University, Nadiad, India.

Abstract- Histone deacetylase inhibitors have been dynamically exposed in epigenetic changes, they mark the genetic processes covering the cell cycle, apoptosis, DNA repair, cell cycle control, autophagy, metabolism, senescence and chaperone function. Several families of histone deacetylase (HDAC) inhibitors have been synthesized and evaluated with their optimistic effects on the cell cycle that have been confirmed in biological models and in clinical trials. Vorinostat, Romidepsin and Belinostat are recently approved for oncologic indications of refractory cutaneous and peripheral T cell Lymphoma. These advances have delivered the stimulus to develop more potent and selective inhibitors and target other pathologic conditions with these drugs. To provide an overview of the use of HDAC inhibitors in cancer treatment, this review addresses the following subjects: (1) the physiological relevance of HDAC-mediated acetylation of histone and nonhistone substrates, and (2) the protein acetylation-independent effect of HDAC inhibitors on the activation status of signaling kinases. (3) Initial and recent development of histone deacetylase inhibitors and chemical classification of inhibitors.

Keywords : Cancer, Histone deacetylase enzyme, Histone deacetylase inhibitors, anticancer therapy,

I. INTRODUCTION

The learning in phenotype without the parallel variation in genotype is characterized as Epigenetics that contains changes in heritability of phenotype from a specific cell to an entity. A group of significant players in transcriptional regulation are two sets of enzymes known as histone acetyl transferases (HATs) and histone deacetylases (HDACs) that controls the N-ε-lysine acetylation and deacetylation of histone. [1-2] Histone deacetylase inhibitors may initiate or inactivate or modify the activities of biological cell cycle [3-4]. Therefore histone deacetylase inhibitors are the main targets for treating human disorders such as cancer, parasitic and inflammatory diseases. The level of histone tail acetylation which regulates the chromatin state is the result of the competing whereabouts of histone acetylase and histone deacetylase enzymes. Histone deacetylase inhibitors change this balance by inhibiting that enzyme, thereby resulting in increased histone acetylation. Histone acetylase and histone deacetylase two classes of enzyme work in opposing directions either by the transfer of acetyl group from acetyl Co A with the help of histone acetylase (HATs) or removing acetyl group with histone deacetylase (HDACs) from lysine residue of histone tails. [5,6] Disturbance of Histone acetylase transferase (HAT) and Histone deacetylase (HDACs) activities has been connected with the increase of a varied range of human cancers. Histone deacetylase (HDACs) inhibitors cause a rise of the acetylated level of histones, which in turn motivates the re-expression of silenced controlling genes in cancer cells and counter the malignant phenotype. Outstanding to this influence, Histone deacetylase (HDACs) inhibitors have recently emerged as prospective cancer therapeutic agents.

II. PHYSIOLOGICAL RELEVANCE OF HDAC-MEDIATED ACETYLATION OF HISTONE AND NONHISTONE SUBSTRATES.

The elementary unit of chromatin is the nucleosome, which comprises of 147 base pairs of DNA superhelix enclosed around the histone core consisting of two copies each of core histones. Histones are the principal protein constituents of chromatin of five classes (H1, H2A, H2B, H3 and H4). H1 is a linker histone and the remaining are core histones. The core plays an important role in establishing interactions between the nucleosomes and within the nucleosome particle itself. The N terminals of core histones are flexible and unstructured, but the rest are predominantly globular and well structured. Chromatin adopts varied conformational changes which subject to epigenetic alterations in DNA and in histone tails that device initiation or suppression of gene transcription. There is on-going of eight distinct histone post transcriptional modifications which are acetylation, methylation, phosphorylation, ubiquitylation, sumoylation, ADP-ribosylation, deamination and proline isomerization.

In eukaryotic cells, DNA wraps around histone proteins, creating complexes called nucleosomes that are filled inside the nucleus. How firmly the negatively charged DNA is wrapped around histones rests on the acetylation state on histone lysine residue, principally on histone H3 and H4. [7,8]

HATs add an acetyl group from acetyl co A to the lysine amino group which reduces the positive charge on the histones. When histone are acetylated, the DNA is more loosely wrapped leading to gene activation. HDACs on the

other hand cause deacetylation ,resulting in positively charged histones that are more tightly wrapped by DNA.Histone deacetylation is correlated with gene repression.The mechanism of action of HDAC inhibitors seems to be multifaceted.HDACs inhibitors are thought to reduce the deacetylation stages of histone proteins that are over communicated in cancerous cells.HAT/HDAC enzymes are capable of regulating both histone and non-histone proteins.Therefore they are capable to exert their therapeutic activity through both histone and non-histone pathways.[9,10]

III. THE PROTEIN ACETYLATION-INDEPENDENT EFFECT OF HDAC INHIBITORS ON THE ACTIVATION STATUS OF SIGNALING KINASES.

Inhibitors of HDACs enzymes are recognized to induce cell cycle arrest, p53 independent induction of cyclin dependent kinase inhibitor p21,tumour selective apoptosis and differentiation of normal and malignant cells.So ,this direct and indirect effect of HDAC enzyme on tumour cells and metastasis make histone deacetylase inhibitors (HDACi)as potential class of anticancer agents. 11,12There are two main pathways of apoptosis, first “extrinsic pathway” or death receptor pathway and second is “intrinsic pathway” or mitochondrial pathway.All HDAC inhibitors have been described to follow either one or both of these cell death pathways in many cancer models.The proposed tools of cancer cell death resulting from HDAC inhibitor treatment are as following:

- I. Death receptor (Extrinsic)pathway of apoptosis
- II.Mitochondrial (Intrinsic)pathway of apoptosis
- III.Inhibition of angiogenesis
- IV.Generation of reactive oxygen species
- V.Autophagy, etc.

Intrinsic pathways are triggered by disruption of mitochondrial membranes in cellular stresses such as chemotherapy , ionizing radiation and withdrawal of growth factors and Extrinsic pathway is originated by binding of ligands,such as Fas ligand(Fas L), tumor necrosis factor (TNF) and TNF –related apoptosis inducing ligand(TRAIL) to their respective cell surface death receptors(DR).

Histone deacetylase inhibitor can also block tumor angiogenesis by reserving of hypoxia inducible factors(HIF)apparatus.Hypoxia controls gene appearance of Vascular Endothelial Growth Factor (VEGF) by improving the transcription factor HIF1 α where as tumor suppressor gene Von Hippel Lindua(VHL)damages HIF1 α .They disrupt heat shock protein 90 (Hsp90) mediated chaperone functionand expose HIF 1 α to proteosomal degradation which contribute to the anti-angiogenic pathway. HDAC inhibitors indirectly damage DNA by promising changes in chromatin conformation upon histone acetylation .It might deduce the DNA to UV rays,ionizing radiation,reactive oxygen species and chemotherapeutic genotoxic chemicals.[13-15]

Generation of reactive oxygen species(ROS) is another main event in this inhibitor to induced cell death,causing DNA damage.Free radical scavengers like N-acetyl cysteine reduce generation of reactive oxygen species which leads to cell death. 16,17]

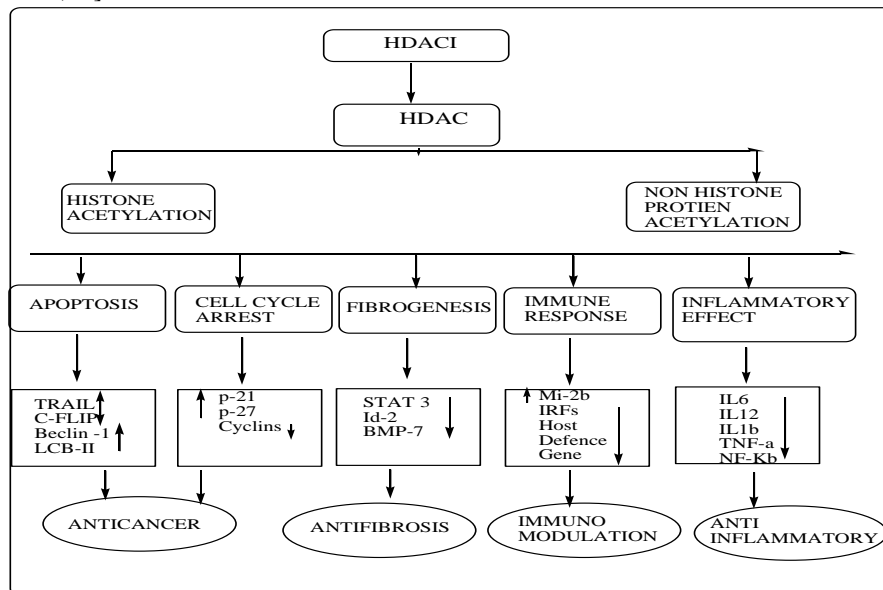


FIGURE 1.Mechanism Of Histone Deacetylase Enzyme

IV. INITIAL DEVELOPMENT OF HDAC IN CLINICAL USE OF CANCER

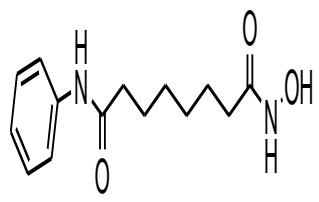
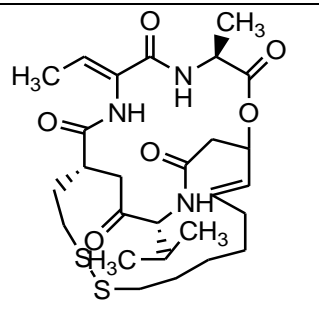
There are four classes of HDAC on their sequence homology to *Saccharomyces cerevisiae* HDACs.[18,19] Eighteen distinct human HDACs are grouped in this four main category. The HDAC family is divided into Zn dependent (Class I and Class II) and NAD –Dependent (Class III) enzymes. The Zn-dependent enzymes have been the focus of strong research, while the Sir2 family recently connected in acetylation and regulation of key cell cycle proteins such as p53. Till date, eleven HDAC family members in classes I and II are considered. i.e. HDACs 1,2,3,8 are class I and HDACs 4-7,9,10 are class II, a grouping based on the system similarity. The most recent identified member of HDAC family is HDAC 11 comprising in Class IV.

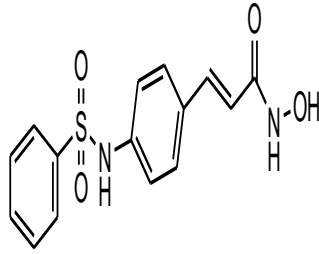
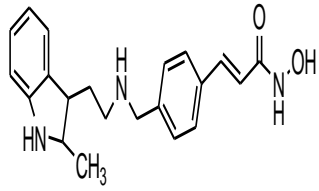
Table 1: Classification of histone deacetylase enzyme

Zn dependant HDAC		NAD-dependant HDAC	
Inhibited by Trichostatin A(TSA) Inhibited by nicotinamide			
Class I	Class II	Class III	
HDAC 1	HDAC 4-7	SIRT 1-7	
HDAC 2	HDAC 9		
HDAC 3	HDAC 10		
HDAC 8			
HDAC 11			

Class I family are homologous to yeast RPD-3 (reduced potassium dependency-3). They share a compact structure. They are predominantly nuclear proteins and ubiquitously expressed in most tissues and cell lines. Class II are homologous to yeast HDA-1-protein (Histone deacetylase-1). It can be subdivided into two subclasses. Class IIa (HDAC 4,5,7,9) have one catalytic domain and long amino terminal adapter domain. Class II b (HDAC 6, HDAC 10) have two catalytic domain. Class II family HDACs are mainly confined in the cytoplasm, however depending upon the phosphorylation status they are split between the cytoplasm and nucleus. Class III inhibitors are also known as sirtuins as they involve of seven members and they share sequence homology with yeast silent information regulator-2 (Sir-2) protein. They do not contain zinc and their activity involve nicotinamide adenine dinucleotide (NAD⁺). Sirtuins (SIRT) are found in three important cellular compartments: nucleus, cytoplasm and mitochondrion. Phylogenetically SIRTs are circulated into four classes (SIRT1, SIRT2 and SIRT3 belong to class I, SIRT4 to class-II, SIRT5 to Class III, and SIRT6 and SIRT7 to class IV.) HDAC 11 has conserved residues in its catalytic center that are shared by both class I and Class II deacetylase and placed in Class IV. [20,21]

Table 2 List Of Clinically Approved Inhibitors

Name	IUPAC Name	Code Name	Trade Name	Chemical Structure	Company/ Approval
Vorinostat (1)	N-Hydroxy-N'-phenyloctane diamide	suberanilohydroxamic acid	Zolinza		Merck (2006) USA
Depsipeptide (2)	1S,4S,7Z,10S,16E,21R)-7-ethylidene-4,21-diisopropyl-2-oxa-12,13-dithia-5,8,20,23-tetraabicyclo[8.7.6]tricos-16-ene-3,6,9,19,22-pentone	Romidepsin	Istodax		Gloucester Pharmaceuticals (2009) USA

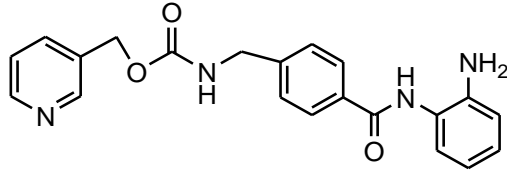
Belinostat (3)	(2E)-N-Hydroxy-3-[3-(phenylsulfamoyl)phenyl]prop-2-enamide	PXD 101	Beleodaq		Spectrum Pharmaceuticals Inc. (2014) USA
Panobinostat (4)	(2E)-N-hydroxy-3-[4-({[2-(2-methyl-1H-indol-3-yl)ethyl]amino}methyl)phenyl]acrylamide	LBH -589	Farydak		Novartis (2015) USA

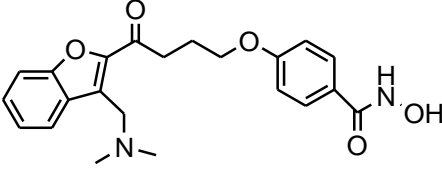
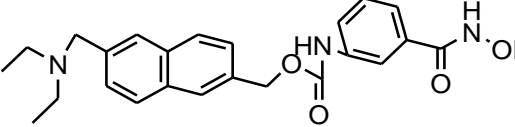
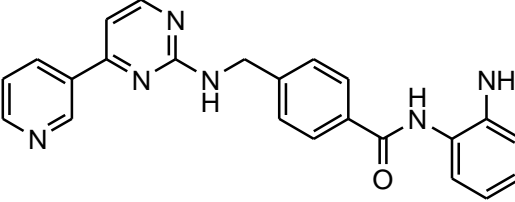
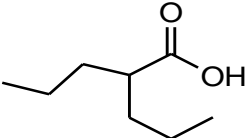
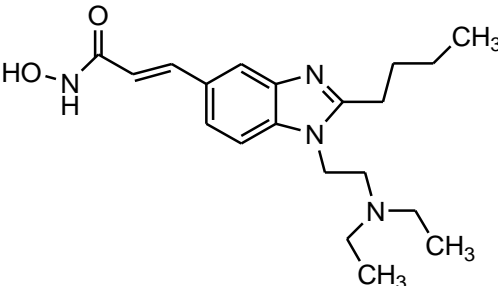
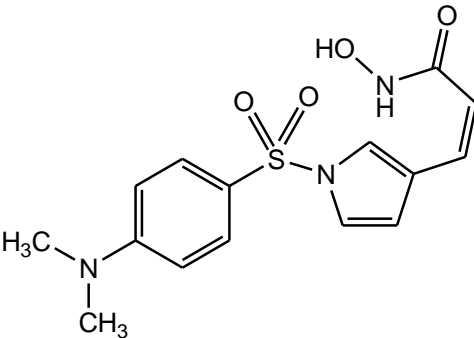
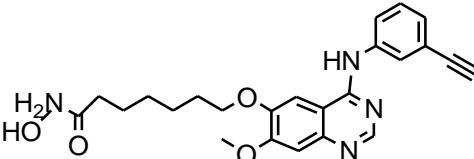
Compound 1 is the first inhibitor used in the treatment of Cutaneous T Cell Lymphoma(CTCL) and sanctioned by Food and drug administration(FDA). After these clinical trials, numerous clinical trials for these inhibitors have been supplemented, causing in the approval and marketing of compound 2 and compound 3 inhibitors. The biological characteristics of these inhibitors differ. Compound 1 is used in phase I and phase II clinical trials for haematological malignancies and solid tumors[22.] Maximum tolerated dose is 400 mg once daily or 200 mg twice daily. Compound 2 is a bicyclic peptide isolated from chromobacterium violaceum and demonstrated potent in vitro cytotoxic activity against human tumor cell lines and in vivo efficacy against human tumor xenograft. It is a unique HDAC inhibitor prodrug. Dose 17.8 mg/m²/day is recommended [23]. In the search for the biologically stable HDAC inhibitors containing benzamide functional group compound 3 was developed. It was investigated in many cell lines which include hepatocellular carcinoma, human cancer, chronic lymphocytic leukemia, prostate cancer, bladder cancer and ovarian cancer cells in [24-30]. Maximum tolerated dose (MTD) is 1000mg/m²/day. Compound 4 is a novel hydroxamate analogue. It demonstrated acetylation of histone H3 and H4 by increasing P21 levels, disrupting chaperone function of hsp90, induce cell cycle G1 phase accumulation and apoptosis of K562 cells and acute leukemia MV4-11 cells.[31]. The anti tumor effect of this drug was demonstrated in multiple myeloma, NSCLC as well as castrate –resistant prostate cancer cell lines. Phase I clinical study with oral compound 4 alone or in combination with docetaxel is also completed. It is being tested against Hodgkin's lymphoma and cutaneous T-cell lymphoma as well as other types of cancer. They can block androgen receptor-mediated transcriptional activation of many genes and thus may result in possible benefit in treating Castration –resistant prostate cancer. [32]

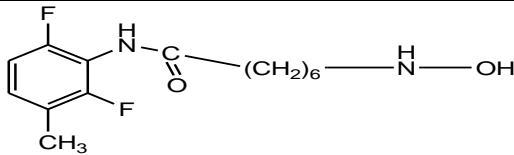
V. MORE RECENT HDAC INHIBITORS UNDER DEVELOPMENT.

The clinical success of the FDA approved drugs following the initial approval of Vorinostat has also stimulated the field to develop and evaluate additional HDAC inhibitors in order to obtain the drugs with improved properties. Several excellent recent reviews have been published covering the development of potential new therapeutic HDAC inhibitors. Several new molecules have reached different phases of clinical trials (TABLE 3) or are in preclinical evaluation.

Table 3 List Of Inhibitors In Ongoing Clinical Studies

Name	Code	Structure	Developer	Clinical Phase	IC50 Value
Entinostat (5)	MS-275, SNDX275		Syndax Pharmaceuticals	II	0.51 μM

Abexinostat (6)	PCI-24781		Pharmacyclins	II	7nM
Givinostat (7)	ITF2357		Investigational Orphan status (2015)	II	7.5nM
Mocetinostat (8)	MGCD-0103		Mirati Therapeutics Inc	II	0.15 μM
Valproic acid (9)	Depakene		Abbott Laboratories	II	5.5nM
Pracinostat (10)	SB939		MEI Pharma Inc	II	40nM
Resminostat (11)	4SC 201		4SC AG	II	42.5 nM
(12)	CUDC-101		Curis ,Inc	II	4.4 nM

(13)	KD 5170		IN preclinical trials	0.054 μM
------	---------	--	-----------------------	-------------

Compound 5 is synthetic benzamide derivative that inhibits HDACi and used to treat patient with leukemia, lymphoma or solid tumors in phase I, II clinical trials. [33] Maximum Tolerated Dose is 10mg/m²/day. Pre-clinical studies suggested that combining inhibitors of DNA methyltransferase (DMT), 5-azacitidine (ADA), with inhibitor of HDAC, SNDX-275, synergistically induced re-expression of epigenetically –silenced tumor suppressor genes and had antitumor effect. Clinical study revealed that it is safe and well tolerated in patients with advanced non small cell lung carcinoma. Dose limiting toxicity were gastrointestinal side effects and fatigue. This class I selective HDAC inhibitor is well accepted either as a single agent or in combination with other drugs.

Compound 6 is formerly CRA 024781. It is phenyl hydroxamic acid which is evaluated alone or with ionizing radiation and other DNA-damaging agents in pre-clinical studies. It acts by preventing DNA repair resulting in synergistic effect on apoptosis when combined with other agents. Phase I clinical trials in refractory advanced solid tumor patients revealed that it was well accepted in following IV or oral route. And it is not dose-related. Mean oral bioavailability was observed to be 0.28%. [34-36] Tubulin and histone acetylation were recognized in peripheral blood mononuclear cells (PBMCs).

Compound 7 is synthesized inhibitor containing a hydroxamic acid linked to aromatic ring. Studies are going on in Phase II clinical study. Many reports have shown that it has inhibitory activity in production of pro-inflammatory cytokines, as well as cytotoxic activity both in-vitro on several human tumor cell lines and in-vivo in patients with hematologic malignancies. [37-41]

Compound 8 is novel isotype selective inhibitor of human HDACi with potential to regulate aberrant gene expression. Phase I/II study with MGCD-0103 alone or combination with genecitabine were done in patients with solid tumors recently. Phase I of clinical trial was studied in refractory solid tumors. Phase II study was done with advanced or metastatic pancreatic cancer. Maximum Tolerated Dose is recommended 90mg, and is ongoing in patients with pancreatic cancer. [42]

Compound 9 is currently used for the treatment of breast cancer, thyroid, lung, ovarian, bladder, head and neck, pancreatic, brain and leukemia cancers. In phase I study, a sequence specific combination of valproic acid and epirubicin in solid tumor malignancies was done. A phase II study of hydralazine and valproic acid in treating patients with advanced solid tumors revealed clinical benefit. Valproic acid shows the good oral bioavailability. 50mg/kg daily dose of valproic acid is given. [43-45]

Compound 10 is in phase II study and tolerated in patients with intermediate or high risk myelofibrosis (MF). This compound is another hydroxamate based inhibitor. This compound was shown to have clinical benefit and modest activity in patients with MF. [46]

Compound 11 is evaluated in a pharmacokinetics and pharmacodynamics phase I study in patients with the advanced solid tumors, yielding a suggested phase II dose of 600 mg/day. Low micro molar concentrations of resminostat lead to cell growth and strongly induced apoptosis in multiple myeloma (MM) cell lines. Synergistic effects were observed with melphalan, bortezomib and S-2209.

Compound 12 has shown that simultaneous inhibition of HDAC and receptor tyrosine kinases (epidermal growth factor receptor-EGFR and human epidermal growth factor receptor-2-HER2) in cancer cells, displayed antiproliferative and proapoptotic activities in vitro as well as in drug –resistant in vivo tumor models. This synergistic inhibition was also tested in patients with advanced solid tumor using this compound, and the drug was found to induce histone H3 acetylation in some of the patients. This study recommended a dose of 275 mg/m² [47]

Compound 13 is a mercaptoketone based class I and II HDAC inhibitor having thioester prodrug demonstrated with broad spectrum cytotoxicity against a range of human tumor-derived cell lines. In the proposed mechanism of action, the thioester prodrug undergoes hydrolysis to generate mercaptoketone that coordinates Zn²⁺ in a bidentate or monodentate fashion in the active site of HDACs. [48]

Histone deacetylase inhibitors are the chemical compounds that inhibit histone deacetylase. A drug is classified into many categories depending on its chemical nature and function. Hence, various different categories of histone deacetylase inhibitors are made.

VI. CHEMICAL STRUCTURE CLASSIFICATION

Most of the available inhibitors possess a three part structure consisting of a zinc binding group that inserts in the active site, linker and a moiety that interact with residues near the entrance to the active site. The classical inhibitors acts entirely on class I and Class II inhibitors by binding to the zinc containing catalytic domain of HDACs. A larger number of natural and synthetic compounds function as inhibitors. Several classes of inhibitors have been identified, including (a) organic hydroxamic acids (b) short chain fatty acids (c) benzamides (d) cyclic tetrapeptides (e) sulphonamide anilides.

6.1 Different Classes Of Hdac Inhibitors

6.1.1. Hydroxamic Acid Derivatives

Hydroxamic acid based derivatives vorinostat is well tolerated in patients with Cutaneous T Cell Lymphoma, and observed promising anti-cancer activities in different types of cancer, such as diffuse large B-cell lymphoma, Hodgkin's lymphoma and other haematological and solid tumors. Given the diverse anticancer activity, much effort has been made to explore hydroxamic acid derivatives such as entinostat, belinostat, givinostat, pracinostat and others as potential treatment for various cancer cells. [49]

6.1.2. Cyclic Peptides

Romidepsin is a bicyclic peptide isolated from the chromobacterium violaceum and demonstrated potent in vitro cytotoxic activity against human tumor cell lines and in vivo efficacy against human tumor xenograft. Natural cyclopeptide FR235222 isolated from the fermentation broth of Acremonium sp. Caused accumulation of acetylated histone H4, inhibition of human leukemia cell (U937) proliferation, and cell cycle arrest in the G1 phase via p21. Other natural cyclopeptides to act as HDAC inhibitors are chlamydocin from *Diheterospora chlamydosporia*, apicidin from *Fusarium* sp. and the microbial metabolite trapoxin. [50]

6.1.3. Benzamide Derivatives

Benzamide containing HDAC inhibitors are another class of compounds that exhibited both in vitro and in vivo anticancer activities. Among them Mocetinostat, Entinostat, Givinostat and Mocetinostat are examples of benzamide derivatives that had been taken into clinical trials. [59]

6.1.4. Short Chain Fatty Acids.

These compounds represent another class of HDAC inhibitor with simple structures that showed clinical potential in various studies. Valproic acid and phenylbutyrate are two well characterized compounds that belong to this class of compounds. They both display inhibition for class I and Class IIa inhibitors. It was proved that hydroxamic acid based HDAC inhibitors are found to be more potent in inhibiting the activity than these drugs. [51]

6.1.5. Miscellaneous

1. Epoxide based histone deacetylase inhibitors

Epoxides are another known group of inhibitors of zinc dependent HDAC enzymes. Epoxide bearing natural compounds such as trapoxin and depudecin are reported to form covalent bonds with HDACs. The HDAC activity of these compounds occur at micro molar to nano molar concentrations. Depudecin is a microbial metabolite containing two epoxide groups, whereas trapoxin has only one epoxide group. [52]

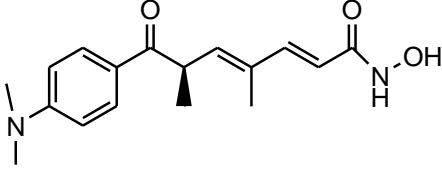
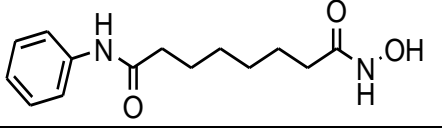
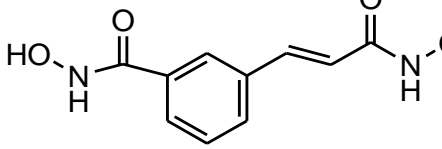
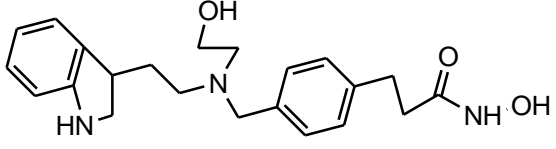
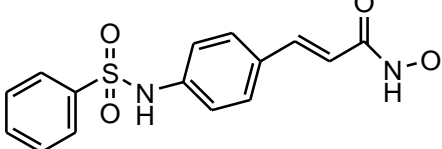
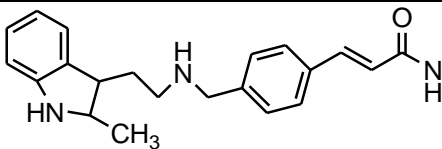
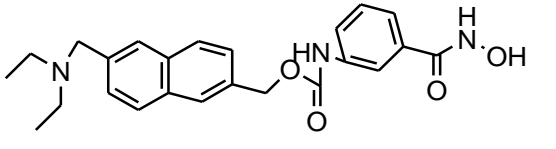
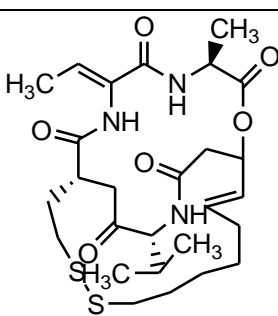
2. Thioester based histone deacetylase inhibitor

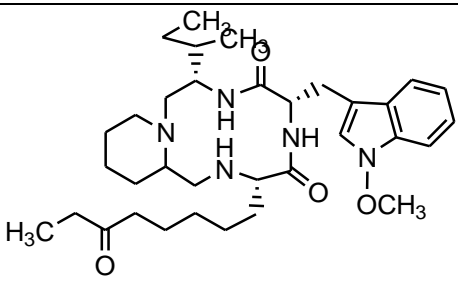
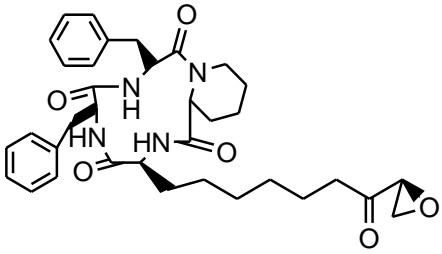
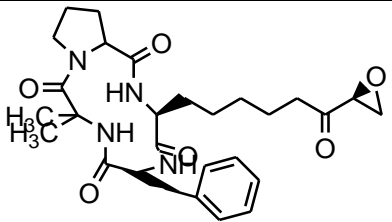
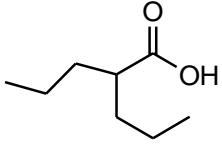
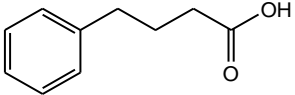
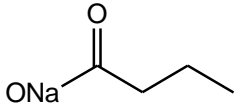
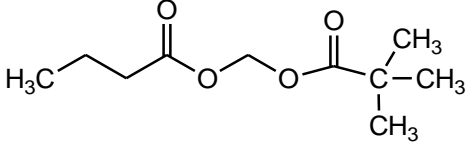
Thioesters are used as prodrug strategies. Largazole is a depsipeptide with the thioester moiety purified from marine cyanobacteria called cyanobacterium *symploea* sp and it is a class I selective HDAC inhibitor. Largazole upon protein assigned hydrolysis liberates the bioactive largazole thiol. Disulfide prodrug strategy to modulate largazole based compounds resulted in enzymatic activities comparable to the natural product largazole.

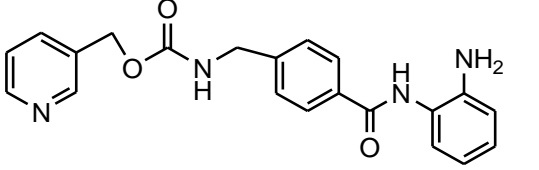
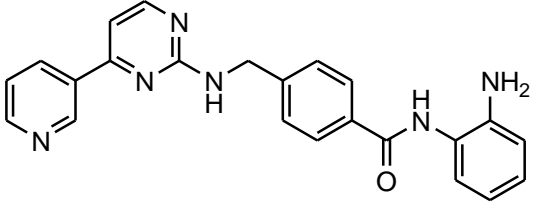
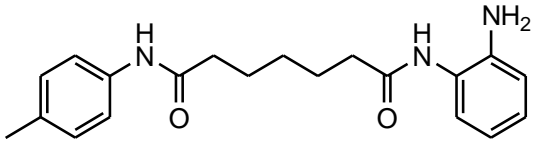
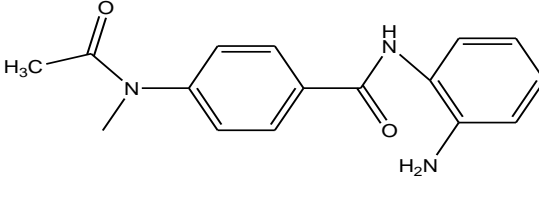
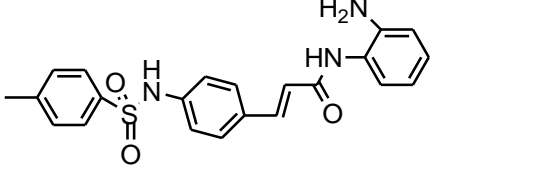
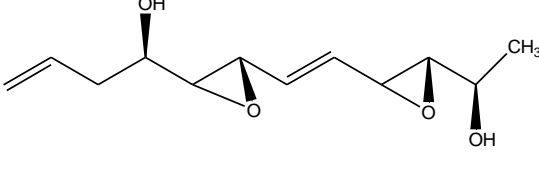
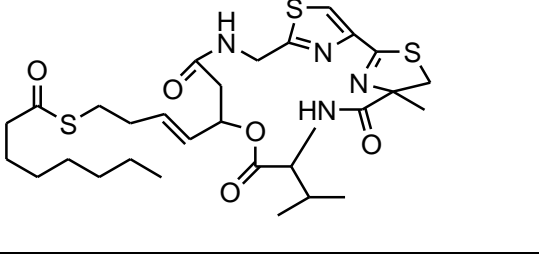
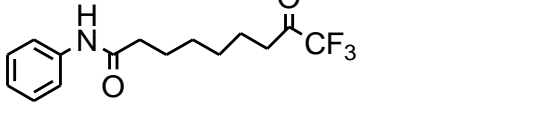
3. Electrophilic ketone based histone deacetylase inhibitor

Trifluoromethyl ketones are known to be readily hydrated and have been reported as potent inhibitors of aspartyl, cysteine and serine proteases, as well as zinc dependent enzymes. Trifluoromethyl ketones attached to aromatic amides showed micro molar inhibitory activities as HDAC inhibitors in breast and fibro sarcoma cell lines. Similarly cyclic tetra peptides containing trifluoromethyl and pentafluoromethyl ketones as zinc binding functional groups were also found to be potent HDAC inhibitors with promising anticancer activities. [53]

Table 4 It shows various structures of HDAC inhibitors along with its detailed chemical classification.

CLASS	COMPOUND	CHEMICAL STRUCTURE
Hydroxamic acid derivatives	TSA(Trichostatin) (14)	
	SAHA(Vorinostat) (1)	
	CBHA (15)	
	LAQ-824(Dacinostat) (16)	
	PDX-101(Belinostat) (3)	
	LBH-589(Panobinostat) (4)	
	ITF-2357(Givinostat) (7)	
Cyclic peptide	Depsipeptide(Romidepsin) (2)	

	Apicidin (17)	
	Trapoxins (18)	
	Chlamydocin (19)	
Short chain fatty acids	Valproic acid (20)	
	Phenylbutyrate (21)	
	Butyrate (22)	
	AN-9 (23)	
Benzamides	Entinostat(MS-275) (5)	

		
	Mocetinostat(MGCD0103) (8)	
	Pimelic diphenylamide (24)	
	N-acetyldinaline (25)	
Sulphonamide Anilides	N-2-aminophenyl-3-[4-(4- methylbenzene sulfonylamino)- phenyl]-2-propenamamide (26)	
Miscellaneous 1.Epoxyde based HDACi	Depudecin (27)	
2.Thioester based HDACi	Largazole (28)	
3.Electrophillic ketone based HDACi	trifluoromethyl ketones (29)	

VII. CONCLUSIONS

Histone deacetylase inhibitors are emerged as recent study of anticancer agents in combination regimens for cancer therapy with the interaction of both the histone and non histone proteins. The development of histone deacetylase inhibitors as anti tumor drugs open a new window in cancer therapeutics. The protagonist of individual histone deacetylase enzyme in tumor development or proliferation as well as therapeutic targets are considered to be essential for tumor studies. The clinical studies have shown that these therapeutic agents cause no or little damage to normal cells and do not show any major side effects. Though vorinostat is active against several malignancy, the reason is not entirely clear. Many other inhibitors are found to be active when clinically tested. Further studies will show whether one representation within the same class or from a different class will be superior to currently approved Vorinostat.

VIII. ABBREVIATIONS

HDACi-Histone deacetylase inhibitor, HAT –histone acetyl transferase, FDA-food and drug administration, MTD-maximum tolerated dose, ZBD-Zinc binding domain, Ac-acetyl group, ROS-reactive oxygen species, VHL- Von Hippel Lindau, CTCL-Cutaneous T cell Lymphoma, DMT-DNA methyl transferases, HIF-Hypoxia inducible factor, VEGF-Vascular endothelial growth factor, Hsp 90- Heat shock protein 90, TSA-Trichostatin A, CDK-cyclin dependent kinase factor.

IX. REFERENCES

- [1] JE Bolden, MJ Peart, RW Johnstone, "Anticancer activities of histone deacetylase inhibitors", *Nat Rev Drug Discov*, 2006,5:pp.769-784
- [2] P Bose, Y Dai, S Grant, "Histone deacetylase inhibitor (HDAC) mechanisms of action: Emerging sights." *Pharmacol Ther*, 2014, pp.358-397.
- [3] Cruz FD, Matushansky I "Solid tumor differentiation therapy-is it possible?" *Oncotarget*, 2012, vol3, pp559-567.
- [4] Dokmanovic M, Clarke C, Marks PA, "Histone deacetylase inhibitors: overview and perspectives." *Mol cancer Res*, 2007,5:981-989.
- [5] Koutsounas I, Giaginis C, Patsouris E, Theocharis S, "Current evidence for histone deacetylase inhibitors in pancreatic cancer." *World J Gastroenterol*, 2013;19:813-828
- [6] Marks PA, Xu WS "Histone deacetylase inhibitors: Potential in cancer therapy" *J cell Biochem*, 2009,107:600-608.
- [7] Xu WS, Parmigiani RB, Marks PA "Histone deacetylase inhibitors: molecular mechanisms of action." *Oncogene*, 2007,26:5541-5552.
- [8] Bieliauskas AV, Pflum MK "Isoform selective histone deacetylase inhibitors." *Chem Soc Rev*, 2008, vol 37:1402-1413.
- [9] West AC, Johnstone RW "New and emerging HDAC inhibitors for cancer treatment." *J Clin Invest*, 2014;124:30-39.
- [10] Zhang L, Han Y, Jiang Q, Wang C, Chen X, et al. "Trend of histone deacetylase inhibitors in cancer therapy: Isoform selectivity or multitargeted strategy." *Med Res Rev*, 2014,3157-3160.
- [11] Kouraklis G, Theocharis S; "Histone deacetylase inhibitors and anticancer therapy." *Curr Med Chem*, 2002, 2:477-484.
- [12] Yang, X.J.; Gregoire, S. Class II histone deacetylases: "From sequence to function, regulation, and clinical implication." *Mol. Cell. Biol.* 2005, 25, 2873–2884.
- [13] Fischle, W.; Kiermer, V.; Dequiedt, F.; Verdin, E. "The emerging role of class II histone deacetylases." *Biochem. Cell Biol.* 2001, 79, 337–348.
- [14] C De Souza -. "HDAC inhibitors as novel cancer therapeutics", *Recent Pat Anticancer Drug Discov*, 2015,145-62
- [15] Lian, Z.R.; Xu, Y.F.; Wang, X.B.; Gong, J.P.; Liu, Z.J. "Suppression of histone deacetylase 11 promotes expression of il-10 in kupffer cells and induces tolerance following orthotopic liver transplantation in rats." *J. Surg. Res.* 2012,174, 359–368.
- [16] Villagra, A.; Cheng, F.; Wang, H.W.; "The histone deacetylase HDAC1 regulates the expression of interleukin 10 and immune tolerance". *Nat. Immunol.* 2009 10, 92–100.
- [17] Buglio, D.; Khaskhely, N.M.; Voo, K.S.; Martinez-Valdez, H.; Liu, Y.J.; Younes, A. "HDAC1 plays an essential role in regulating ox40 ligand expression in hodgkin lymphoma". *Blood*, 2011,117, 2910–2917.
- [18] Hess-Stumpff H, Bracker TU, Henderson D, Politz O "MS-275, a potent orally available inhibitor of histone deacetylases-the development of an anticancer agent". *Int J Biochem Cell Biol.* 2007,39:1388-1405.
- [19] Luca DM, Davis ME, Parthun MR, Mone AP, Kitada S, Cunningham KD et al "The histone deacetylase inhibitor MS-275 induces caspase-dependant apoptosis in B-Cell chronic lymphocytic leukemia cells." *Leukemia*, 2004,18:1207-1214.
- [20] Marks P, Rifkin RA, Richon VM, Breslow R, Miller T, Kelly WK; "Histone deacetylase and cancer: causes and therapies." *Nat Rev Cancer*, 2001, vol1:194-202
- [21] Buggy JJ, Cao ZA, Bass KE, Verner E, Balasubramanian S, Liu L, "CRA-024781: a novel synthetic inhibitor of histone deacetylase enzymes with antitumor activity in vivo and in vitro." *Mol Cancer Ther.* 2006,5:1309-1317.
- [22] Grant S, Easley C, Kirkpatrick P "Vorinostat". *Nat Rev Drug Discov*; 2007;6:21-22.
- [23] Sandor V, Bakke S, Robey RW, Kang MH, Blagosklonny MV, Bender J, et al "Phase I trial of histone deacetylase inhibitor, desipeptide (FR90 NSC 630176), in patients with refractory neoplasms." *Clin Cancer Res*, 2008, vol 12:718-728.
- [24] Marshall JL, Rizvi N, Kauh J, Dahut W, Figueroa M, Kang MH, et al; "A phase I trial of desipeptide in patients with advanced cancer." *J Exp Ther Oncol*, 2002, vol2:325-332.
- [25] Welsbie DS, Xu J, Chen Y, Borsu L, Scher HI, Rosen N "Histone deacetylases are required for androgen receptor function in hormone sensitive and castrate –resistant prostate cancer." *Cancer Res*, 2009,69:958-966.
- [26] Buckley MT, Yoon J, Yee H, Chiriboga L, Liebes L, Ara G; "The histone deacetylase inhibitor belinostat (PXD101) suppresses bladder cancer cell growth in vitro and in vivo." *J Transl Med*, 2007, vol5:49.
- [27] Dai Y, Chen S, Kramer L B, Funk VL, Dent P, Grant S "Interactions between bortezomib and romidepsin and belinostat in chronic lymphocytic leukemia cells." *Clin Cancer Res*, 2008,14:549-558.
- [28] Deljigbjerg M, Gruaslund M, Christensen JJ, Tjoeneland J, Buhl JP, Sehested M, "Identification of predictive biomarkers for the histone deacetylase inhibitor belinostat in a panel of human cancer cell lines." *Cancer Biomark*, 2008,4:101-109.

- [29] Duan J,Friedman J,Nottingham L,Chen Z,Ara G,Van WC “Nuclear factor kappaB p65 small interfering RNA or proteasome inhibitor bortezomib sensitizes head and neck squamous cell carcinoma to classic histone deacetylase inhibitors and novel histone deacetylase inhibitor PXD101. “Mol Cancer Ther,2007,6:37-50.
- [30] Ramalingam SS,Belani CP,Ruel C,Frankel P,Gitlitz B,Koczywas M ”Phase II study of belinostat (PXD101),a histone deacetylase inhibitor,for second line therapy of advanced malignant pleural mesothelioma“.J Thorac Oncol,2009,4:97-101.
- [31] Ma BB,Sung F,Tao Q,Poon FF,Lui –VW,Yeo W. “The preclinical activity of the histone deacetylase inhibitor PXD101(belinostat)in hepatocellular carcinoma cell lines. “Invest New Drugs,2009,100-105.
- [32] Qian X,LaRoche W J,Ara G,Wu F,Petersen KD,Thougaard A, “Activity of PXD101,a histone deacetylase inhibitor,in preclinical ovarian cancer studies.Mol Cancer Ther”,2006,5:2086-2095.
- [33] Qian X,Ara G,Mills E,Larochelle WJ,Linchenstien HS,Jeffers M “Activity of the histone deacetylase inhibitor belinostat(PXD101) in preclinical models of prostate cancer. “Int J Cancer,2008,122:1400-1410.
- [34] Ellis L ,Pan Y,Smyth GK,George DJ,McCormack C,Williams –Truax R,“Histone deacetylase inhibitor panobinostat induces clinical responses with associated alterations in gene expression profiles in cutaneous T- Cell lymphoma. “Clin Cancer Res,2008,14:4500-4510.
- [35] Rosato RR,Almenara JA,Grant S; “The histone deacetylase inhibitor MS-275 promotes differentiation or apoptosis in human leukemia cells through a process regulated by generation of reactive oxygen species and induction of p21 CIP1/WAF1“.Cancer Res,2003,63:3637-3645.
- [36] Banuelos CA,Banath JP,MacPhail SH,Zhao J,Rietsema T,Olive PL; “Radiosensitization by the histone deacetylase inhibitor PCI-24781“.Clin Cancer Res ,2007,13:6816-6826.
- [37] Adimoolam S,Siriaswad M,Chen J,Thiemann P ,Ford JM, Buggy JJ“HDAC inhibitors PCI-24781 decreases RAD51 expression and inhibits homologous recombination. “Proc Natl Acad Sci, 2007,USA 104:19482-19487.
- [38] Undevia SD,Janisch L,Schilsky RL,Loury D,Balasubramanian S,Mani C ; “Phase I study of safety,pharmacokinetics(PK) and pharmacodynamics(PD) of the histone deacetylase inhibitor(HDACi)PCI-24781 “. J Clin Oncol,2008,26,1451
- [39] Barbetti V,Gozzini A,Rovida E,Morandi A,Spinelli E,Fossati G; “Selective anti-leukemic activity of low dose histone deacetylase inhibitor ITF2357 on AML1/ETO-positive cells “.Oncogene,2008,vol 27:1767-1778.
- [40] Petrini M,Galimberti S,Canestraro M,Salvi H,Palumbo GA,Nagy B “Histone deacetylase inhibitor ITF2357 is effective on the P39 cells.A gene expression study “. J Clin Oncol.2008,26(32):5156-5164.
- [41] Salvi H,Galimberti S,Canestraro M,Nagy B,Palumbo GA,Petrini M,“Bortezomib combined with histone deacetylase inhibitor ITF2357 or arsenic trioxide exerts synergistic anti proliferative and pro-apoptotic effect on P39 cells. “.J.Clin Oncol, 2008,26(32):5156-5164.
- [42] Golay J,Cuppini L,Leoni F,Mico C,Barbui V,Domenghini M,et al“The histone deacetylase inhibitor ITF2357 has anti-leukemic activity in vitro and in vivo and inhibits IL-6 and VEGF production by stromal cells “.Leukemia,2007,21:1892-1900.
- [43] Qlian DZ,Kato Y,Shabbeer S,WeiY,Verhuel HM,Salumbides et al: “Targetting tumor angiogenesis with histone deacetylase inhibitors:the hydroxamic acid derivative LBH589“. Clin Cancer Res,2006,12:634-642.
- [44] Geng L,Cuneo KC,Fu A ,Tu T ,Atadja PW ,Hallahan DE; “Histone deacetylase inhibitor(HDAC) LBH589 increases duration of gamma H2AX foci and confines HDAC4 to the cytoplasm in irradiated non small cell lung cancer. “Cancer Res, 2006,66:11298-11304.
- [45] Svetlana, H.; Tanja, P.; Stefan, H.ITOC2 – 031” Immunomodulatory characteristics of Resminostat, a novel HDAC inhibitor in phase II clinical development “. European Journal of Cancer.2015,Vol 51, , pp. S11
- [46] Voi, M.; Fu, S.; Nemunaitis, J.; Bauman, J.; Bessudo, A.; Hamid, O.; Witta, S.; Dy, G.; Lai, C.; Leliberte, R.590 ”Final Results of a Phase Ib Study of CUDC-101, a Multitargeted Inhibitor of EGFR, HER2, and HDAC, in Patients with Advanced Head and Neck, Gastric, Breast, Liver, and Non-small Cell Lung Cancer “.European Journal of Cancer,2012, Vol 48, 6, pp. 181
- [47] Hassig CA,Symons KT, Guo X, Nguyen PM, Annable T. “KD5170, a novel mercaptoketone-based histone deacetylase inhibitor that exhibits broad spectrum antitumor activity in vitro and in vivo.Molecular CancerTherapy,2008,150-157.
- [48] Bali P,Pranpat M,Bradner J,Balasis M,Fisku W,Guo F,et al,Inhibition of histone deacetylase 6 acetylates and disrupts the chaperone function of heat shock protein 90:a novel basis for antileukemia activity of histone deacetylase inhibitors.J Biol Chem,2005,280:26729-26734.
- [49] Haefner M,Bluethner T,Nielderhagen M,Moebius C,Wittekind C,Mossner J,et al,“Experimental treatment of pancreatic cancer with two novel histone deacetylase inhibitors.” World J Gastroenterol,2008,14:3681-3692.
- [50] Kim JC,Kim DD,Lee YM,Kim TW,Cho DH,Kim MB,et al,“Evaluation of novel histone deacetylase inhibitors as therapeutic agents for colorectal adenocarcinomas compared to established regimens with the histoculture drug response array “.Int J Colorectal Dis,2009,24:209-218.
- [51] Meeran S M,Patel S N,Li Y,Shukla S,Tollefsbol TO “Bioactive dietary supplements reactivate ER expression in ER-negative breast cancer cells by active chromatin modifications: “PLoS one ,2012,7:37748.
- [52] Bangert A,Christofanon S,Eckhardt I,Akbari BA,Kolodziej S. “Histone deacetylase inhibitors sensitize glioblastoma cells to TRAIL – induced apoptosis by c-myc-mediated downregulation of cFLIP “.Oncogene,2012,31:4677-4688.
- [53] Goldberg AD,Allis CD,Bernstein E, ”Epigenetics :a landscape takes shape “.Cell,2007, 128:635-638.